REVIEW

Open Access

Cancer predictive studies



Ivano Amelio^{1*}, Riccardo Bertolo^{1,2}, Pierluigi Bove^{1,2}, Eleonora Candi¹, Marcello Chiocchi¹, Chiara Cipriani^{1,2}, Nicola Di Daniele¹, Carlo Ganini¹, Hartmut Juhl³, Alessandro Mauriello¹, Carla Marani^{1,2}, John Marshall⁴, Manuela Montanaro¹, Giampiero Palmieri¹, Mauro Piacentini¹, Giuseppe Sica¹, Manfredi Tesauro¹, Valentina Rovella¹, Giuseppe Tisone¹, Yufang Shi^{1,5,6}, Ying Wang⁵ and Gerry Melino^{1*}

Abstract

The identification of individual or clusters of predictive genetic alterations might help in defining the outcome of cancer treatment, allowing for the stratification of patients into distinct cohorts for selective therapeutic protocols. Neuroblastoma (NB) is the most common extracranial childhood tumour, clinically defined in five distinct stages (1–4 & 4S), where stages 3–4 define chemotherapy-resistant, highly aggressive disease phases. NB is a model for geneticists and molecular biologists to classify genetic abnormalities and identify causative disease genes. Despite highly intensive basic research, improvements on clinical outcome have been predominantly observed for less aggressive cancers, that is stages 1,2 and 4S. Therefore, stages 3–4 NB are still complicated at the therapeutic level and require more intense fundamental research. Using neuroblastoma as a model system, here we herein outline how cancer prediction studies can help at steering preclinical and clinical research toward the identification and exploitation of specific genetic landscape. This might result in maximising the therapeutic success and minimizing harmful effects in cancer patients.

Keywords: Neuroblastoma, Microbiota, Precision oncology, Omics

Background

Since the revelation of the whole human genome, there has been tremendous advances in sequencing technologies, with reductions of costs and time, allowing for an incredible step forward in the global cancer genomic fruition [1–3]. We moved from The Cancer Genome Atlas (TCGA) to the Pan-Cancer Analysis of Whole Genomes (PCAWG) Consortium [4–6], which includes most tumor types, matching DNA sequencing and RNA transcripts. Recently, 2658 cancers have been deeply analysed [7, 8] reconstructing the origin and evolution of mutational processes and driver mutation sequences of 38 types of

* Correspondence: ivano.amelio@uniroma2.it; melino@uniroma2.it; melino@uniroma2.it

¹Torvergata Oncoscience Research Centre of Excellence, TOR, Department of Experimental Medicine, University of Rome Tor Vergata, via Montpellier 1, 00133 Rome, Italy

Full list of author information is available at the end of the article



Since the clinical application of massive sequencing, and moreover the clinical application of omics, each patient can provide an enormous amount of molecular data which can be also implemented in the drug discovery process, moving fast towards the selection of the right drug for the right patient. This attitude towards the introduction of precision medicine into clinical therapy, was furthermore boosted by President Obama's speech introducing the Precision Medicine Initiative in 2015, aimed at creating a collaborative academical environment [9, 10] to improve the use of the omics in both patients' treatments and drug development.

Alongside with the advances in sequencing technologies, drug discovery rapidly reshaped itself. Drugs were



© The Author(s). 2020 **Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit http://creativecommons.org/licenses/by/4.0/. The Creative Commons Public Domain Dedication waiver (http://creativecommons.org/publicdomain/zero/1.0/) applies to the data made available in this article, unless otherwise stated in a credit line to the data.

initially discovered in a serendipitous fashion as in the case of penicillin [11], or aspirin [12]. Pharmaceutical efforts were aimed at finding molecules suitable for a given unmet medical need, as in the case of infections or inflammatory diseases. A single agent was tested in a single disease at a time. Chemical libraries speeded up the process, allowing the testing of multiple molecules in a relatively cheap and fast approach, and leading to the discovery of many classes of drugs which now have a predominant role in the clinics, such as statins [13]. At this point, the scientific focus was still on the molecule itself and the pathology for which it was developed. Patients' clinical characteristics were still not fully evaluated during the drug development process, due to the yet inadequate availability of patient-derived molecular data. Nevertheless, further advances in the technologies used in this disease-oriented drug discovery approach, such as fragment screening of chemical compounds [14, 15] and lately cryo-electron microscopy [16] or structural bioinformatics [17], led to the development of a large number of therapies. This abundance of molecules, however, does not reflect the higher complexity of many diseases, among which cancer represents a prominent example: in many clinical contexts there is an abundance of therapies which can be chosen, but in many cases patients would not respond primarily [18]. Understanding the molecular profile of each of those primary refractory patients might represent a possible way to overcome our inability to tailor the right treatment for the right patient. This is in desperate need for primary refractory patients, as well as for patients who would benefit from many of the existing treatments but without knowing which one would get to the maximum clinical efficacy. Figure 1 reports a brief outline of the evolution of therapeutic drug development.

Massive sequencing approaches applied to disease entities such as cancer, have changed our point of view on diseases themselves, initially considered as entities per se, to disease as a single individual related clinical condition; this paradigm shift allowed to move to a completely patient-oriented effort to select the right drug, at the right moment. Effectively, under this light, many clinical trials have undergone a revolution: clinical efficacy is not anymore the only primary objective since also the study of biomarkers, to define subpopulations of treated or to be treated patients who would benefit more from a given treatment, is now performed [19, 20]. The vast availability of molecular data from patients, derived from multiple omics approaches, together with the implementation of computing algorithms and artificial intelligence, also opened a new era in drug discovery and clinical practice [21-23]: the selection of the right molecular target can be done on a single patient basis, and its efficacy and potential toxicity can also be predicted using a metabolomic approach in combination with genetic information [24].

Precision medicine has therefore entered the clinic in a single patient perspective [25–27]. This is an extremely actual approach, especially in disease entities which have been extensively characterized from a molecular point of view in the era of global sequencing approaches such as Neuroblastoma.

The case of neuroblastoma

Neuroblastoma (NB) is clinically linked to distinct genomic abnormalities that seems to involve possible causative and progression genes of the disease [28–30]. These genomic abnormalities include deletions on chromosomes 1p, 11q or gains on 17q2,3; all being effective prognostic markers of the clinical outcome even though their molecular mechanisms are still not fully elucidated. Another crucial NB genetic signature is the amplification of the proto-oncogene MYCN; its amplification, with its overexpression, is a stronger predictor of tumor aggressiveness, chemotherapy refractoriness and clinical outcome. For example, NB patients with MYCN amplification report less than 50% 5 years survival, whilst the non-MYCN-amplified might report over 90%.

Nevertheless, MYCN is not the only culprit of NB aggressiveness. Recently, familial or sporadic NB patients have been reported to carry activating mutations of ALK. In the former cases, up to 50% of the cases show germline mutations in ALK gene, while sporadic NB may acquire ALK somatic mutations and ~ 2% display genomic amplification. ALK, member of the receptor tyrosine kinases superfamily and in particular, the insulin receptor (IR), shows homology with the leukocyte tyrosine kinase, the insulin-like growth factor-1 receptor kinase and the IR kinase. The single-chain transmembrane ALK is localized on human chromosome 2p23. At the molecular level, its mutation/amplification fosters cell proliferation and survival via the JAK-STAT, PI3K-AKT or RAS-MAPK pathways. Indeed, mutated (constitutively activated) ALK physically binds hyperphosphorylated ShcC, inhibiting, in response to growth factors, the MAPK signalling. Moreover, NB shows the deletion or loss-of-function mutation of the RNA-helicase ATRX. Interestingly, deregulation of ATRX and MYCN are mutually exclusive.

A further unexpected genetic rearrangement in high-risk NB is the activation of telomerase reverse transcriptase (TERT). This occurs in the chromosomal region 5p15.33 proximal of TERT. Again, TERT rearrangements, ATRX mutations and MYCN amplifications are mutually exclusive even though they take place exclusively within the high-risk NB patients. This leads to the concept that these genetic abnormalities converge on a similar function. Still, in MYCN-amplified tumors without TERT rearrangements, its expression is nevertheless increased: juxtapose TERT rearrangements to strong enhancers result in deep epigenetic remodelling of the regulatory region without changes in the



genetic banks, with specific mutations, deletions, polymorphisms and histone modifications (phase 3). This permitted the identification of intelligent drugs, acting only on one single target and therefore wanton toxicity (phase 4), in other wards, selecting the specific drug for the individual patient. With metabolic mutation, identification of predisposing mutations, selection of monitoring or predictive cluster of genes, proteins or phospho-protein, oncology will enter the 4P medicine: Preventive, Personalized, Predictive, Participative

gene copy number. Whole-genomic sequencing shows that ATRX mutations occur only in MYCN-non-amplified and TERT-normal NB and are associated with increase in alternative lengthening of telomeres (ALT). This indicates that telomere lengthening is a common trait of high-risk NB (MYCN-amplified, ATRX-mutated, TERT-rearranged cancers) independently from the underlying molecular mechanism of telomere maintenance. Therefore, high-risk NB show telomerase activation that is subsequent to either TERT rearrangement or MYCN amplification (which in any case is able to activates TERT).

The Chr17q region containing the TRIM37 gene is frequently amplified in neuroblastoma, as well as breast cancer. Since the acentrosomal spindle assembly following PLK4 inhibition, during mitotic division, depends on levels of the centrosomal ubiquitin ligase TRIM37 [31]. The steady state level of TRIM37 regulates the spindle assembly and subsequently the proliferation, in particular following PLK4 inhibition. Therefore, TRIM37 is a prognostic factor for human NB with 17p-deletion and, at the same time, an essential determinant of mitotic vulnerability to PLK4 inhibition [31]. This is highly relevant, as recently excellent PLK4 inhibitors have been identified [32, 33]. A PLK4 inhibitor, CFI-400945, triggers mitotic catastrophe in breast cancer cells overexpressing TRIM37 [34].

The locus deleted in 1p in NB contains an interesting gene, Trp73, codifying for the p73 protein [35, 36]. p53, p63 and p73 define the p53 family of transcription factors. All three are transcribed as several distinct protein

isoforms [37-40]. Two alternative promoters drive the expression of either a transcriptionally active p73 (TA isoforms) proteins [41], containing a full N-terminal transactivation domain (TAD), or a N-terminally truncated (ΔN isoforms) proteins [42], that lack the TAD. ΔN isoforms might have fully independent functions or may act as dominant negative molecules by inhibiting the transactivating activity of the TA isoforms. For example, while TAp73 is an inducer of cell cycle arrest, neuronal function [43-45] and apoptotic cell death, and largely mimics the tumor suppressive activities of p53 [46–53], $\Delta Np73$ isoforms promote cancer cell survival and exhibit oncogenic properties. The phenotypical characterization of selectively knockout mice for either TAp73 and Δ Np73 fully support their function as tumor suppressor or pro-oncogenic factors, respectively. Moreover, p73 is essential for the development and differentiation of the neuronal tissue. Accordingly, TAp73^{-/-} knockout mice as well as p73^{D13/D13} knockout mice show hippocampal dysgenesis with reduction of the neurogenesis in the subgranular zone of the dentate gyrus [54]. Conversely, $\Delta Np73^{-/-}$ knockout mice exhibit signs of neurodegeneration, as a consequence of the prosurvival function of this isoforms. Hence, TAp73 and $\Delta Np73$ are crucial regulators of tumorigenesis and neurodevelopment [55, 56].

Recently, we discovered that expression of ZNF281, a zinc finger factor associated with several cellular functions, is deregulated in terminal differentiation of murine cortical neurons and in differentiating NB cells. Indeed, the mouse zinc finger transcription factor Zfp281 (or the human homologue ZNF281), involved in the control of neuronal progenitor stemness by inhibiting Nanog expression in mice through recruitment of the inhibitory complex NuRD on the Nanog promoter, is significantly expressed in neuronal cells, and significantly repressed during neuronal differentiation, including neuroblastoma [57]. ZNF281 is highly expressed in stage 4 NB patients supporting a role of ZNF281 in the progression of the disease. Accordingly, NB patients with "low-expressors", thus indicating that ZNF281 represents a prognostic marker of human NB [57].

Another member of the same family is also expressed in NB tumours: ZNF143 [58]. In this case, LIN28B, or LIN28B mutant that is unable to inhibit let-7 processing, increases the penetrance of MYCNinduced neuroblastoma, potentiates the invasion and migration of transformed sympathetic neuroblasts, and drives distant metastases in vivo. In particular, LIN28B physically binds ZNF143 and activates the expression of downstream targets, that is GSK3B and L1CAM, affecting adhesion and migration of the cancer cells [58]. Figure 2 shows how the distinct NB subgroups with distinct genetic aberrations define individual prognostic groups.

It is therefore essential, following a diagnosis of NB, to understand which molecular defect is present in each individual patient, in order characterize the patient's risk group and select the more appropriate therapy, accordingly. In each of these groups, the identification of the





underlying molecular events at the bases of tumour progression [59–61] might allow for specific combination therapies to optimise therapeutic efficacy and minimise toxic side effects [62].

Is there a role for exogenous factors such as microbiome?

The microbiome [63–66], often interacting with environmental urban life [67–70], constitutes a large mass of metabolizing bacteria, which are able to metabolise and transform normal constituents [71–73] and to impinge on the function of the host [74–77]. Just as examples, microbiota directly affects the B cell repertoire [78], the histone HDAC3 activity [79] or even the function of mutant p53 [80, 81]. Importantly, microbiota can also affect immunity to tumors and the efficacy of chemotherapies, but can also affect massively inflammatory cronic diseases [82, 83].

This, just to remain with the neuroblastoma example, are also of relevance in other cancer progression or even in neurophysiology. Accordingly, gut microbiota shows neuroprotective properties, reducing IL-6 secretion in different neural cell lines, including neuroblastoma [84, 85]. Here, these investigators identified two specific strains, Parabacteroides distasonis MRx0005 and Megasphaera massiliensis MRx0029, producing distinct C1-C3 or C4-C6 fatty acids [84] or, in another context, galacto-oligosaccharides [86] or short-chain fatty acid receptor 3 [87]. Similarly, Roseovarius albus increases brain derived neurotropic factor (BDNF) expression while reducing Bax/Bcl-2 ratio in neuroblastoma cell lines [88]. Neuroblastoma has per se the ability to impinge on the gut microbiota [89]. Consequently, it is conceivable to provide supplementary dietary treatment in neuroblastoma patients to improve the therapeutic response [90–93].

Conclusion

The advance in technology, including massive sequencing, bioinformatic analysis by artificial intelligence, cloud computing, fast large scale proteomic and phospho-proteomic analysis is rapidly providing a unique opportunity to the global cancer genomic community to improve the previous analysis of TCGA with an individual systematic documentation of selective mutations which drive common tumour types. This provides, as being done now, the use of intelligent drugs with a single target for an individual patient, therefore reducing undesirable toxicity, while increasing the efficacy. In the near future, such possibilities will be expanded along with the use of the 4P medicine: Preventive, Personalized, Predictive, Participative.

Acknowledgements

Not applicable.

Authors' contributions

IA, GM and CG wrote the manuscript. GM prepared the figures. All the other indicated authors (RB, PB, EC, MC, CC, NDD, HJ, AM, CM, JM, MM, GP, MP, GS, MT, VR, GT, YS, YW) made substantial contribution to the conception of the manuscript and critically revised it. All of the Authors have approved this submitted version.

Funding

This work has been supported by the Associazione Italiana per la Ricerca contro il Cancro (AIRC) to GM (IG#20473; 2018–2022), to IA (AIRC Start-Up ID 23219; 2020–2024), to EC (IG#22206; 2019–2023), Ministry of Health & MAECI Italy-China Science and Technology Cooperation (#PGR00961) to GM and YW.

Availability of data and materials

Not applicable.

Ethics approval and consent to participate

Not applicable.

Consent for publication

Not applicable.

Competing interests

The Authors declare that they have no competing interests.

Author details

¹Torvergata Oncoscience Research Centre of Excellence, TOR, Department of Experimental Medicine, University of Rome Tor Vergata, via Montpellier 1, 00133 Rome, Italy. ²San Carlo di Nancy Hospital, Rome, Italy. ³Indivumed GmbH, Hamburg, Germany. ⁴Medstar Georgetown University Hospital, Georgetown University, Washington DC, USA. ⁵CAS Key Laboratory of Tissue Microenvironment and Tumor, Shanghai Institute of Nutrition and Health, Shanghai Institutes for Biological Sciences, University of Chinese Academy of Sciences, Chinese Academy of Sciences, 320 Yueyang Road, Shanghai 200031, China. ⁶The First Affiliated Hospital of Soochow University and State Key Laboratory of Radiation Medicine and Protection, Institutes for Translational Medicine, Soochow University, 199 Renai Road, Suzhou 215123, Jiangsu, China.

Received: 28 September 2020 Accepted: 2 October 2020 Published online: 14 October 2020

References

- Kim SY, Jeong H-H, Kim J, Moon J-H, Sohn K-A. Robust pathway-based multi-omics data integration using directed random walks for survival prediction in multiple cancer studies. Biol Direct. 2019;14(1):8.
- Panchin AY, Aleoshin W, Panchin YV. From tumors to species: a SCANDAL hypothesis. Biol Direct. 2019;14(1):3.
- Qu Q, Li Y, Fang X, Zhang L, Xue C, Ge X, et al. Differentially expressed tRFs in CD5 positive relapsed & amp; refractory diffuse large B cell lymphoma and the bioinformatic analysis for their potential clinical use. Biol Direct. 2019;14(1):23.
- Pcawg Transcriptome Core Group C, Calabrese C, Davidson NR, Demircioğlu D, Fonseca NA, He Y, et al. Genomic basis for RNA alterations in cancer. Nature. 2020;578(7793):129–36.
- Alexandrov LB, Kim J, Haradhvala NJ, Huang MN, Tian Ng AW, Wu Y, et al. The repertoire of mutational signatures in human cancer. Nature. 2020; 578(7793):94–101.
- Mihaylov I, Kańduła M, Krachunov M, Vassilev D. A novel framework for horizontal and vertical data integration in cancer studies with application to survival time prediction models. Biol Direct. 2019;14(1):22.
- Gerstung M, Jolly C, Leshchiner I, Dentro SC. The evolutionary history of 2 ,658 cancers. Nature. 2020;578(August 2017).
- Rheinbay E, Nielsen MM, Abascal F, Wala JA, Shapira O, Tiao G, et al. Analyses of non-coding somatic drivers in 2,658 cancer whole genomes. Nature. 2020;578(7793):102–11.
- Terry SF. Obama's precision medicine initiative. Genet Test Mol Biomarkers. 2015;19(3):113–4.
- Nathan C. Cooperative development of antimicrobials: looking back to look ahead. Nat Rev Microbiol. 2015;13(10):651–7.

- 11. Leisner JJ. The diverse search for synthetic, semisynthetic and natural product antibiotics from the 1940s and up to 1960 exemplified by a small pharmaceutical player. Front Microbiol. 2020;11:976.
- Desborough MJR, Keeling DM. The aspirin story from willow to wonder drug. Br J Haematol. 2017;177(5):674–83.
- Endo A. A historical perspective on the discovery of statins. Proc Jpn Acad Ser B Phys Biol Sci. 2010;86(5):484–93.
- Gerry CJ, Schreiber SL. Chemical probes and drug leads from advances in synthetic planning and methodology. Nat Rev Drug Discov. 2018;17(5):333–52.
- 15. Vajda S, Whitty A, Kozakov D. Fragments and hot spots in drug discovery. Oncotarget. 2015;6(22):18740–1.
- Scapin G, Potter CS, Carragher B. Cryo-EM for small molecules discovery, design, understanding, and application. Cell Chem Biol. 2018;25(11):1318–25.
- Gaudreault F, Morency LP, Najmanovich RJ. NRGsuite: a PyMOL plugin to perform docking simulations in real time using FlexAID. Bioinformatics. 2015;31(23):3856–8.
- Massari F, Di Nunno V, Santoni M, Gatto L, Caserta C, Morelli F, et al. Toward a genome-based treatment landscape for renal cell carcinoma. Crit Rev Oncol Hematol. 2019;142:141–52.
- Polley MC, Korn EL, Freidlin B. Phase III precision medicine clinical trial designs that integrate treatment and biomarker evaluation. JCO Precis Oncol. 2019;3.
- Insabato L, Amelio I, Quarto M, Zannetti A, Tolino F, de Mauro G, et al. Elevated expression of the tyrosine phosphatase SHP-1 defines a subset of high-grade breast tumors. Oncology. 2009;77(6):378–84.
- 21. Fleming N. How artificial intelligence is changing drug discovery. Nature. 2018;557(7707):S55–s7.
- Chen B, Garmire L, Calvisi DF, Chua MS, Kelley RK, Chen X. Harnessing big 'omics' data and Al for drug discovery in hepatocellular carcinoma. Nat Rev Gastroenterol Hepatol. 2020;17(4):238–51.
- Wolfender JL, Litaudon M, Touboul D, Queiroz EF. Innovative omics-based approaches for prioritisation and targeted isolation of natural products new strategies for drug discovery. Nat Prod Rep. 2019;36(6):855–68.
- 24. Wishart DS. Emerging applications of metabolomics in drug discovery and precision medicine. Nat Rev Drug Discov. 2016;15(7):473–84.
- Jameson JL, Longo DL. Precision medicine–personalized, problematic, and promising. N Engl J Med. 2015;372(23):2229–34.
- Prasad V. Perspective: the precision-oncology illusion. Nature. 2016; 537(7619):S63.
- 27. Abrahams E, Eck SL. Molecular medicine: Precision oncology is not an illusion. Nature. 2016;539:357.
- Nicolai S, Pieraccioli M, Peschiaroli A, Melino G, Raschellà G. Neuroblastoma: oncogenic mechanisms and therapeutic exploitation of necroptosis. Cell Death Dis. 2015;6(12):e2010.
- Polewko-Klim A, Lesiński W, Mnich K, Piliszek R, Rudnicki WR. Integration of multiple types of genetic markers for neuroblastoma may contribute to improved prediction of the overall survival. Biol Direct. 2018;13(1):17.
- Suo C, Deng W, Vu TN, Li M, Shi L, Pawitan Y. Accumulation of potential driver genes with genomic alterations predicts survival of high-risk neuroblastoma patients. Biol Direct. 2018;13(1):14.
- Meitinger F, Ohta M, Lee K-Y, Watanabe S, Davis RL, Anzola JV, et al. TRIM37 controls cancer-specific vulnerability to PLK4 inhibition. Nature. 2020; 585(7825):440–6.
- Press MF, Xie B, Davenport S, Zhou Y, Guzman R, Nolan GP, et al. Role for polo-like kinase 4 in mediation of cytokinesis. Proc Natl Acad Sci U S A. 2019;116(23):11309–18.
- Kawakami M, Mustachio LM, Zheng L, Chen Y, Rodriguez-Canales J, Mino B, et al. Polo-like kinase 4 inhibition produces polyploidy and apoptotic death of lung cancers. Proc Natl Acad Sci U S A. 2018;115(8):1913–8.
- Yeow ZY, Lambrus BG, Marlow R, Zhan KH, Durin MA, Evans LT, et al. Targeting TRIM37-driven centrosome dysfunction in 17q23-amplified breast cancer. Nature. 2020;585(7825):447–52.
- Kaghad M, Bonnet H, Yang A, Creancier L, Biscan JC, Valent A, et al. Monoallelically expressed gene related to p53 at 1p36, a region frequently deleted in neuroblastoma and other human cancers. Cell. 1997;90(4):809–19.
- Levrero M, De Laurenzi V, Costanzo A, Gong J, Melino G, Wang JY. Structure, function and regulation of p63 and p73. Cell Death Differ. 1999;6(12):1146–53.
- Amelio I, Mancini M, Petrova V, Cairns RA, Vikhreva P, Nicolai S, et al. p53 mutants cooperate with HIF-1 in transcriptional regulation of extracellular matrix components to promote tumor progression. Proc Natl Acad Sci U S A. 2018;115(46):E10869–E78.

- Amelio I, Inoue S, Markert EK, Levine AJ, Knight RA, Mak TW, et al. TAp73 opposes tumor angiogenesis by promoting hypoxia-inducible factor 1α degradation. Proc Natl Acad Sci U S A. 2015;112(1):226–31.
- 39. Amelio I, Cutruzzolá F, Antonov A, Agostini M, Melino G. Serine and glycine metabolism in cancer. Trends Biochem Sci. 2014;39(4):191–8.
- Ho CJ, Lin RW, Zhu WH, Wen TK, Hu CJ, Lee YL, et al. Transcriptionindependent and -dependent p53-mediated apoptosis in response to genotoxic and non-genotoxic stress. Cell Death Dis. 2019;5:131.
- Tomasini R, Tsuchihara K, Wilhelm M, Fujitani M, Rufini A, Cheung CC, et al. TAp73 knockout shows genomic instability with infertility and tumor suppressor functions. Genes Dev. 2008;22(19):2677–91.
- Wilhelm MT, Rufini A, Wetzel MK, Tsuchihara K, Inoue S, Tomasini R, et al. Isoform-specific p73 knockout mice reveal a novel role for delta Np73 in the DNA damage response pathway. Genes Dev. 2010;24(6):549–60.
- Agostini M, Tucci P, Killick R, Candi E, Sayan BS. Rivetti di Val Cervo P, et al. neuronal differentiation by TAp73 is mediated by microRNA-34a regulation of synaptic protein targets. Proc Natl Acad Sci U S A. 2011;108(52):21093–8.
- Billon N, Terrinoni A, Jolicoeur C, McCarthy A, Richardson WD, Melino G, et al. Roles for p53 and p73 during oligodendrocyte development. Development. 2004;131(6):1211–20.
- Rada M, Barlev N, Macip S. BTK modulates p73 activity to induce apoptosis independently of p53. Cell Death Dis. 2018;4:30.
- Bernassola F, Salomoni P, Oberst A, Di Como CJ, Pagano M, Melino G, et al. Ubiquitin-dependent degradation of p73 is inhibited by PML. J Exp Med. 2004;199(11):1545–57.
- Melino G, Knight RA, Nicotera P. How many ways to die? How many different models of cell death? Cell Death Differ. 2005;12(Suppl 2):1457–62.
- Nemajerova A, Amelio I, Gebel J, Dotsch V, Melino G, Moll UM. Nononcogenic roles of TAp73: from multiciliogenesis to metabolism. Cell Death Differ. 2018;25(1):144–53.
- Amelio I, Antonov AA, Catani MV, Massoud R, Bernassola F, Knight RA, et al. TAp73 promotes anabolism. Oncotarget. 2014;5(24):12820–934.
- Celardo I, Antonov A, Amelio I, Annicchiarico-Petruzzelli M, Melino G. p63 transcriptionally regulates the expression of matrix metallopeptidase 13. Oncotarget. 2014;5(5):1279–89.
- Amelio I. How mutant p53 empowers Foxh1 fostering leukaemogenesis? Cell Death Dis. 2019;5:108.
- McSweeney KM, Bozza WP, Alterovitz WL, Zhang B. Transcriptomic profiling reveals p53 as a key regulator of doxorubicin-induced cardiotoxicity. Cell Death Dis. 2019;5:102.
- Amelio I, Melino G. The p53 family and the hypoxia-inducible factors (HIFs): determinants of cancer progression. Trends Biochem Sci. 2015;40(8):425–34.
- Amelio I, Panatta E, Niklison-Chirou MV, Steinert JR, Agostini M, Morone N, et al. The C terminus of p73 is essential for hippocampal development. Proc Natl Acad Sci U S A. 2020;117(27):15694–701.
- Agostini M, Romeo F, Inoue S, Niklison-Chirou MV, Elia AJ, Dinsdale D, et al. Metabolic reprogramming during neuronal differentiation. Cell Death Differ. 2016;23(9):1502–14.
- Hooper C, Meimaridou E, Tavassoli M, Melino G, Lovestone S, Killick R. p53 is upregulated in Alzheimer's disease and induces tau phosphorylation in HEK293a cells. Neurosci Lett. 2007;418(1):34–7.
- Pieraccioli M, Nicolai S, Pitolli C, Agostini M, Antonov A, Malewicz M, et al. ZNF281 inhibits neuronal differentiation and is a prognostic marker for neuroblastoma. Proc Natl Acad Sci U S A. 2018;115(28):7356–61.
- Tao T, Shi H, Mariani L, Abraham BJ, Durbin AD, Zimmerman MW, et al. LIN28B regulates transcription and potentiates MYCN-induced neuroblastoma through binding to ZNF143 at target gene promotors. Proc Natl Acad Sci U S A. 2020;117(28):16516–26.
- Grimes T, Walker AR, Datta S, Datta S. Predicting survival times for neuroblastoma patients using RNA-seq expression profiles. Biol Direct. 2018; 13(1):11.
- Hidalgo MR, Amadoz A, Çubuk C, Carbonell-Caballero J, Dopazo J. Models of cell signaling uncover molecular mechanisms of high-risk neuroblastoma and predict disease outcome. Biol Direct. 2018;13(1):16.
- Amelio I, Tsvetkov PO, Knight RA, Lisitsa A, Melino G, Antonov AV. SynTarget: an online tool to test the synergetic effect of genes on survival outcome in cancer. Cell Death Differ. 2016;23(5):912.
- 62. Tranchevent L-C, Nazarov PV, Kaoma T, Schmartz GP, Muller A, Kim S-Y, et al. Predicting clinical outcome of neuroblastoma patients using an integrative network-based approach. Biol Direct. 2018;13(1):12.

- 63. Matute JD, Duan J, Blumberg RS. Microbial RNAs Pressure Piezo1 to Respond. Cell. 2020;182(3):542–4.
- 64. Osmanovic D, Kessler DA, Rabin Y, Soen Y. Darwinian selection of host and bacteria supports emergence of Lamarckian-like adaptation of the system as a whole. Biol Direct. 2018;13(1):24.
- Marafini I, Monteleone I, Di Fusco D, Cupi ML, Paoluzi OA, Colantoni A, et al. TNF-α producing innate lymphoid cells (ILCs) are increased in active celiac disease and contribute to promote intestinal atrophy in mice. PLoS One. 2015;10(5):e0126291.
- Biancone L, Onali S, Calabrese E, Petruzziello C, Zorzi F, Condino G, et al. Non-invasive techniques for assessing postoperative recurrence in Crohn's disease. Dig Liver Dis. 2008;40(Suppl 2):S265–70.
- Walker AR, Grimes TL, Datta S, Datta S. Unraveling bacterial fingerprints of city subways from microbiome 16S gene profiles. Biol Direct. 2018;13(1):10.
- Zolfo M, Asnicar F, Manghi P, Pasolli E, Tett A, Segata N. Profiling microbial strains in urban environments using metagenomic sequencing data. Biol Direct. 2018;13(1):9.
- Zhu C, Miller M, Lusskin N, Mahlich Y, Wang Y, Zeng Z, et al. Fingerprinting cities: differentiating subway microbiome functionality. Biol Direct. 2019; 14(1):19.
- Casimiro-Soriguer CS, Loucera C, Perez Florido J, López-López D, Dopazo J. Antibiotic resistance and metabolic profiles as functional biomarkers that accurately predict the geographic origin of city metagenomics samples. Biol Direct. 2019;14(1):15.
- Kawulok J, Kawulok M, Deorowicz S. Environmental metagenome classification for constructing a microbiome fingerprint. Biol Direct. 2019; 14(1):20.
- Walker AR, Datta S. Identification of city specific important bacterial signature for the MetaSUB CAMDA challenge microbiome data. Biol Direct. 2019;14(1):11.
- Caputo A, Fournier P-E, Raoult D. Genome and pan-genome analysis to classify emerging bacteria. Biol Direct. 2019;14(1):5.
- Tuganbaev T, Mor U, Bashiardes S, Liwinski T, Nobs SP, Leshem A, et al. Diet Diurnally Regulates Small Intestinal Microbiome-Epithelial-Immune Homeostasis and Enteritis. Cell. 2020;182(6):1441–59.
- 75. Gerner SM, Rattei T, Graf AB. Assessment of urban microbiome assemblies with the help of targeted in silico gold standards. Biol Direct. 2018;13(1):22.
- 76. Ryan FJ. Application of machine learning techniques for creating urban microbial fingerprints. Biol Direct. 2019;14(1):13.
- Mars RAT, Yang Y, Ward T, Houtti M, Priya S, Lekatz HR, et al. Longitudinal Multi-omics Reveals Subset-Specific Mechanisms Underlying Irritable Bowel Syndrome. Cell. 2020;182(6):1460–73.
- Li H, Limenitakis JP, Greiff V, Yilmaz B, Schären O, Urbaniak C, et al. Mucosal or systemic microbiota exposures shape the B cell repertoire. Nature. 2020; 584(7820):274–8.
- Wu S-E, Hashimoto-Hill S, Woo V, Eshleman EM, Whitt J, Engleman L, et al. Microbiota-derived metabolite promotes HDAC3 activity in the gut. Nature. 2020;586(7827):108–12.
- Kadosh E, Snir-Alkalay I, Venkatachalam A, May S, Lasry A, Elyada E, et al. The gut microbiome switches mutant p53 from tumour-suppressive to oncogenic. Nature. 2020;586(7827):133–8.
- Bulatov E, Sayarova R, Mingaleeva R, Miftakhova R, Gomzikova M, Ignatyev Y, et al. Isatin-Schiff base-copper (II) complex induces cell death in p53positive tumors. Cell Death Dis. 2018;4:103.
- Sileri P, Sica G, Gentileschi P, Venza M, Manzelli A, Palmieri G, et al. Ischemic preconditioning protects intestine from prolonged ischemia. Transplant Proc. 2004;36(2):283–5.
- Onali S, Calabrese E, Petruzziello C, Zorzi F, Sica GS, Lolli E, et al. Endoscopic vs ultrasonographic findings related to Crohn's disease recurrence: a prospective longitudinal study at 3 years. J Crohns Colitis. 2010;4(3):319–28.
- Ahmed S, Busetti A, Fotiadou P, Vincy Jose N, Reid S, Georgieva M, et al. In vitro Characterization of Gut Microbiota-Derived Bacterial Strains With Neuroprotective Properties. Front Cell Neurosci. 2019;13:402.
- Garrido-Maraver J, Celardo I, Costa AC, Lehmann S, Loh SHY, Martins LM. Enhancing folic acid metabolism suppresses defects associated with loss of Drosophila mitofusin. Cell Death Dis. 2019;10(4):288.
- Savignac HM, Corona G, Mills H, Chen L, Spencer JPE, Tzortzis G, et al. Prebiotic feeding elevates central brain derived neurotrophic factor, Nmethyl-D-aspartate receptor subunits and D-serine. Neurochem Int. 2013; 63(8):756–64.

- Getachew B, Csoka AB, Bhatti A, Copeland RL, Tizabi Y. Butyrate protects against Salsolinol-induced toxicity in SH-SY5Y cells: implication for Parkinson's disease. Neurotox Res. 2020;38(3):596–602.
- Park J, Lee J, Yeom Z, Heo D, Lim Y-H. Neuroprotective effect of Ruminococcus albus on oxidatively stressed SH-SY5Y cells and animals. Sci Rep. 2017;7(1):14520.
- Castellani C, Singer G, Kaiser M, Kaiser T, Huang J, Sperl D, et al. Neuroblastoma causes alterations of the intestinal microbiome, gut hormones, inflammatory cytokines, and bile acid composition. Pediatr Blood Cancer. 2017;64(8).
- Jang H-M, Lee K-E, Kim D-H. The Preventive and Curative Effects of Lactobacillus reuteri NK33 and Bifidobacterium adolescentis NK98 on Immobilization Stress-Induced Anxiety/Depression and Colitis in Mice. Nutrients. 2019;11(4)..
- Castellani C, Singer G, Eibisberger M, Obermüller B, Warncke G, Miekisch W, et al. The effects of neuroblastoma and chemotherapy on metabolism, fecal microbiome, volatile organic compounds, and gut barrier function in a murine model. Pediatr Res. 2019;85(4):546–55.
- Obermüller B, Singer G, Kienesberger B, Klymiuk I, Sperl D, Stadlbauer V, et al. The Effects of Prebiotic Supplementation with OMNI-LOGIC® FIBRE on Fecal Microbiome, Fecal Volatile Organic Compounds, and Gut Permeability in Murine Neuroblastoma-Induced Tumor-Associated Cachexia. Nutrients. 2020;12(7):2029.
- Gite S, Ross RP, Kirke D, Guihéneuf F, Aussant J, Stengel DB, et al. Nutraceuticals to promote neuronal plasticity in response to corticosteroneinduced stress in human neuroblastoma cells. Nutr Neurosci. 2019;22(8): 551–68.

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Ready to submit your research? Choose BMC and benefit from:

- fast, convenient online submission
- · thorough peer review by experienced researchers in your field
- rapid publication on acceptance
- support for research data, including large and complex data types
- gold Open Access which fosters wider collaboration and increased citations
- maximum visibility for your research: over 100M website views per year

At BMC, research is always in progress.

Learn more biomedcentral.com/submissions

